

DENEOSAMINYLLIVIDOMYCIN B

TAKASHI YAMAGUCHI, KAZUHIRO KAMIYA,
TOSHIHITO MORI and TAKESHI ODATokyo Research Laboratories, Kowa Co., Ltd.
Higashimurayama, Tokyo 189, Japan

(Received for publication February 8, 1977)

In our previous papers,¹⁻⁵⁾ the structural elucidation of lividomycins A and B isolated from the fermentation broth of *Streptomyces lividus* nov. sp., and the chemical conversion of lividomycin A into lividomycin B by elimination of the mannose moiety were reported. In the present paper, we report a chemical conversion of lividomycin B into a pseudotrisaccharide by elimination of the neosamine B moiety according to the method reported by HANESSIAN⁶⁾. The pseudotrisaccharide named deneosaminyllividomycin B is weakly bioactive, but useful as a starting material for the synthesis of therapeutically important aminoglycoside antibiotics, such as 3'-deoxyribostamycin and 3'-deoxybutirosin B.

An aqueous solution (50 ml) of penta-N-acetyllividomycin B⁵⁾ (I, 2.0 g) was treated with sodium periodate (0.8 g) in the dark at room temperature for 20 hours with stirring. After addition of ethylene glycol (0.3 ml), the reaction mixture was evaporated to dryness and the residue (2.8 g) was extracted with methanol (30 ml). The dialdehyde derivative (II) in the methanol solution was detected by thin-layer chromatography on Silica gel 60F (E. Merck) using

chloroform - methanol (1:1 by volume), visualized by triphenyltetrazolium chloride reagent as a single spot with R_f 0.50. Treatment of II in the methanol solution with 3 ml of triethylamine at 25°C for 4 hours, followed by evaporation afforded tri-N-acetyldeneosaminyllividomycin B (III, 1.12 g, 78%), which was purified by column chromatography (2.0 × 27.0 cm) on Silica gel C-200 (Wako Chemicals, 40 g) eluted with chloroform - methanol (3:2 by volume), mp. 173~176°C (dec.), [α]_D²⁰ +49° (c 0.5, methanol). Anal. calcd. for C₂₃H₃₉N₅O₁₃·1/2H₂O: C, 48.08; H, 7.02; N, 7.31%. Found: C, 48.06; H, 7.01; N, 7.41%.

A solution of III (900 mg) in 4 N sodium hydroxide (22 ml) was heated on a boiling water bath for 5 hours. The reaction mixture was neutralized with 4 N hydrochloric acid, diluted with water (200 ml) and passed through a column (2.0 × 17.0 cm) of Amberlite IRC-50(NH₄⁺) resin. After washing the column with water (300 ml), elution with 0.3 N aqueous ammonia (200 ml) gave a ninhydrin-positive effluent. Rechromatography on a column (2.0 × 25.0 cm) of CM-Sephadex C-25(NH₄⁺) using a gradient elution between water (300 ml) and 0.2 N aqueous ammonia (300 ml) afforded deneosaminyllividomycin B (IV) as a white powder (580 mg) in 83% yield. By thin-layer chromatography on Silica gel 60F (E. Merck) using chloroform - methanol - 17% aqueous ammonia (1:4:3 by volume), IV gave a single spot with R_{lividomycin B} 1.24 after detection with the ninhydrin reagent. By addi-

Scheme 1.

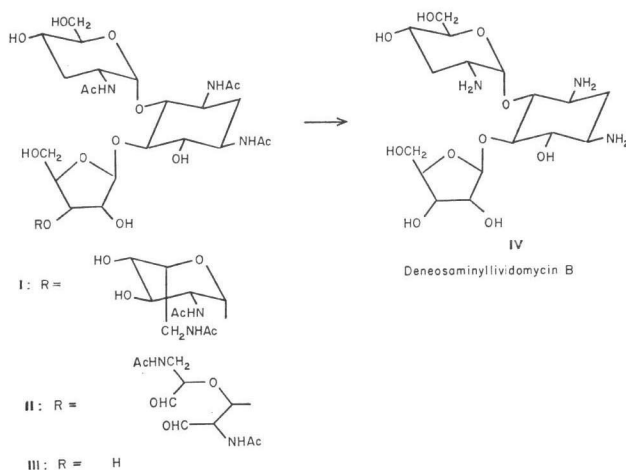
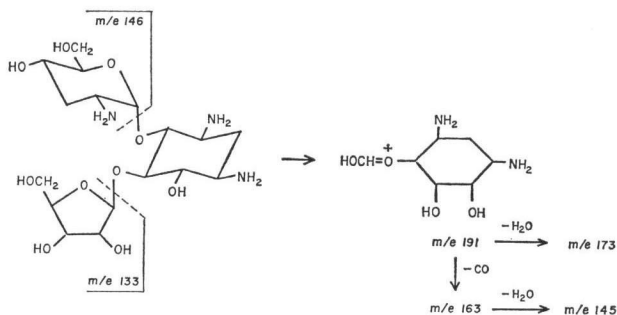


Fig. 1. Significant fragments in mass spectrum of deneosaminyllividomycin B



tion of an equimolar amount of H_2SO_4 to an aqueous solution of IV (100 mg), the sulfate was obtained as a white powder (141 mg), mp. $170\sim 174^\circ\text{C}$ (dec.), $[\alpha]_D^{25} +30^\circ$ (c 0.8, water). Anal. calcd. for $\text{C}_{17}\text{H}_{33}\text{N}_3\text{O}_{10}\cdot 3/2\text{H}_2\text{SO}_4\cdot 2\text{H}_2\text{O}$: C, 32.80; H, 6.47; N, 6.75; S, 7.72%. Found: C, 32.67; H, 6.10; N, 6.91; S, 8.02%.

After methanolysis of IV with 0.4 N hydrogen chloride in methanol at 28°C for 6 days, only 3'-deoxyparomamine and methyl riboside were detected by thin-layer chromatography. The structure of IV was further confirmed by the direct electron impact mass spectrum as shown in Fig. 1. Both m/e 440 $[(M+1)^+]$ and m/e 439 (M^+) peaks were sufficiently abundant for clear assignment of molecular composition. The intensive series of peaks at m/e 191, 173, 163 and 145 indicated a 2-deoxystreptomine-containing compound⁷⁾. A very intensive peak at m/e 146 was assigned to the 3'-deoxyglucosamine unit, which was produced by normal glycosidic cleavage. On the other hand, the peak of the ribofuranosyl unit (m/e 133) was rather weak⁷⁾.

Acknowledgements

The authors wish to acknowledge Dr. H. UMEZAWA and Dr. S. KONDO, Institute of Microbial Chemistry, for their helpful advice and encouragement in the performance of this work. Thanks are also due to Prof. M. OHASHI, University of Electro-communications, for the measurement of mass spectra and valuable discussions.

References

- 1) ODA, T.; T. MORI, H. ITÔ, T. KUNIEDA & K. MUNAKATA: Studies on new antibiotic lividomycins. I. Taxonomic studies on the lividomycin-producing strain *Streptomyces lividus* nov. sp. *J. Antibiotics* 24: 333~338, 1971
- 2) MORI, T.; T. ICHIYANAGI, H. KONDÔ, K. TOKUNAGA, T. ODA & K. MUNAKATA: Studies on new antibiotic lividomycins. II. Isolation and characterization of lividomycins A, B and other aminoglycosidic antibiotics produced by *Streptomyces lividus*. *J. Antibiotics* 24: 339~346, 1971
- 3) ODA, T.; T. MORI & Y. KYÔTANI: Studies on new antibiotic lividomycins. III. Partial structure of lividomycin A. *J. Antibiotics* 24: 503~510, 1971
- 4) ODA, T.; T. MORI, Y. KYÔTANI & M. NAKAYAMA: Studies on new antibiotic lividomycins. IV. Structure of lividomycin A. *J. Antibiotics* 24: 511~518, 1971
- 5) MORI, T.; Y. KYÔTANI, I. WATANABE & T. ODA: Chemical conversion of lividomycin A into lividomycin B. *J. Antibiotics* 25: 149~150, 1972
- 6) TAKAMOTO, T. & S. HANESSIAN: Aminoglycoside antibiotics. Chemical transformation of paromomycin into a bioactive pseudotrisaccharide. *Tetrahedron Letters* 1974-46: 4009~4012, 1974
- 7) DANIELS, P. J. L.; A. K. MALLAMS, J. WEINSTEIN, J. J. WRIGHT & G. W. A. MINE: Mass spectral studies on aminocyclitol-aminoglycoside antibiotics. *J. Chem. Soc. Perkin I*: 1976: 1078~1088, 1976